



---

# Polarimetric interferometry to objectively evaluate the optical properties of corneal stroma

Eugenio Lipari<sup>1</sup>, Alessandra Sborgia<sup>2</sup>, Mario Nubile<sup>3</sup>, Leonardo Mastropasqua<sup>3</sup>, Giovanni Alessio<sup>2</sup>

<sup>1</sup>Phronema SRL, Bari, Italy; <sup>2</sup>Department of Medical Basic Sciences, Neuroscience and Sense Organs, Ophthalmology Clinic, University of Bari "A. Moro", Bari, Italy; <sup>3</sup>Department of Medicine and Ageing Sciences, Ophthalmology Clinic, University of Chieti-Pescara "G. d'Annunzio", Chieti, Italy

## Abstract

A new non-invasive method, based on the interferometric analysis of diffractive and polarizing effects related to the birefringent properties of corneal collagen fibrils, has been developed to objectively evaluate the optical properties of the stroma. The new method shows a relevant impact on corneal surgeries specifically for lamellar transplantation where, due to the polarizing properties of the stroma, the alignment between collagen fibrils of donor corneas with patient collagen fibril orientation has shown an improvement of visual acuity postoperatively. Further studies on the regularity of the corneal isogyre pattern are showing this new method has a strong impact in early-stage diagnosis of corneal disease.

*Keywords:* corneal cross, isogyre, melatope, birefringence

## 1. Introduction

The cornea has a dual function, optical and mechanical. The human cornea provides two-thirds of the refractive power of the eye and the requisite durability to maintain its shape, notwithstanding the action of the extraocular muscles and the internal force of the ciliary muscle. The latter mediates the change in lens shape in order

---

**Correspondence:** Eugenio Lipari, Phronema SRL, Via Junipero Serra n. 19, Bari, Italy.  
E-mail: lipari@phronema.it

---

to adjust the focusing power of the eye, and has been shown to have a very small and optically negligible effect on corneal shape.<sup>1</sup> A fundamental characteristic for corneal function is that of transparency. The cornea is classically accepted as a five-layered structure: an epithelial and an endothelial layer at the anterior and posterior surfaces respectively; two membranes: Bowman's under the epithelium and Descemet's anterior to the endothelium; and a central and predominant stroma composed of collagen fibrils, which represent approximately 90% of corneal thickness.

The stroma is composed of approximately two successively stacked lamellae of type I collagen fibrils with a diameter of around 25-30  $\mu\text{m}$ .<sup>2</sup> Within each lamella, the collagen fibrils run parallel to each other and show a regular interfibrillar spacing.<sup>2-5</sup> The orientation of the fibrils is constant within each lamella, but varies throughout successive layers.<sup>6-9</sup> The regular arrangement of the fibrils within each layer is considered to be responsible for the transparency of the tissue.<sup>2,10,11</sup> It is also an important factor for determining the mechanical properties of the cornea<sup>12,13</sup> as well as maintaining its shape.<sup>14,15</sup>

Such an organization contributes to characteristic patterns that are linked to highly ordered structures, such as crystals, which possess the property of birefringence.

These characteristic patterns seen when the structure is illuminated between crossed polarizers are a dark cross, the arms of which have been called isogyres, and colored rings or isochromatics. Whilst these formations, which are caused by the separation of refracted rays into ordinary and extraordinary components that travel at different velocities through the crystal depending on the atomic spacing in the plane of travel, can be explained in relation to crystal structure,<sup>16</sup> similar features have been noted over 200 years ago in eye lenses.<sup>17-18</sup> More recently, they have been noted in the cornea.<sup>7,8,19-24</sup> This cannot be explained on the same basis as crystals, given that ocular tissue is not highly ordered, but rather accords with the descriptions for elongated structures that arise in biology, such as cell layer arrangements.<sup>25,26</sup> The lamellar organization of the corneal stroma is akin to a Wiener body,<sup>27</sup> which gives rise to form birefringence due to the directional variations in refractive index: *i.e.*, differences along single fibers and across fiber layers.<sup>27</sup> It has also been shown that isogyres can be formed in curved structures made of amorphous materials that do not possess any birefringent properties.<sup>21,28</sup> The number of studies that have considered corneal birefringence show differences in findings,<sup>19-24</sup> and indeed it has been suggested that the random orientation of the central layers of the stroma results in no birefringence effects at the corneal apex.<sup>8</sup> Such variation is to be expected in a biological tissue that differs in shape and thickness between individuals and changes with age and pathologies. There has been a paucity of investigation into the use of polarization optics for the study of stromal structure. Stromal orientation is important for optical quality and differences between individuals may suggest that the particular orientation of lamellae is optimized

for image quality. In cases of corneal transplantation, the orientation of the donor cornea should therefore be considered. This study presents results of measurements on *in-vivo* corneal tissue using a new device that can determine the polarization properties of the cornea *in vitro* or *in vivo*. The instrument applies specialized software to determine the orientation of the corneal fibrils, offering the prospect of a detailed structural analysis which may have applications for clinical studies.

## 2. Scope

Development of a new non-invasive method to objectively evaluate the optical properties of the stroma, based on the interferometric analysis of diffractive and polarizing effects related to the birefringent properties of the stroma.

## 3. Method

Comparison between polarimetric interferometry image obtained by the interference between polarized light and stromal structure of human corneas using a new patented<sup>29</sup> medical device called Lumaxis® (Phronema SRL; Bari, Italy) with data published in the literature and obtained by x-ray, and second and third harmonic generation technique (SGH eTGH).

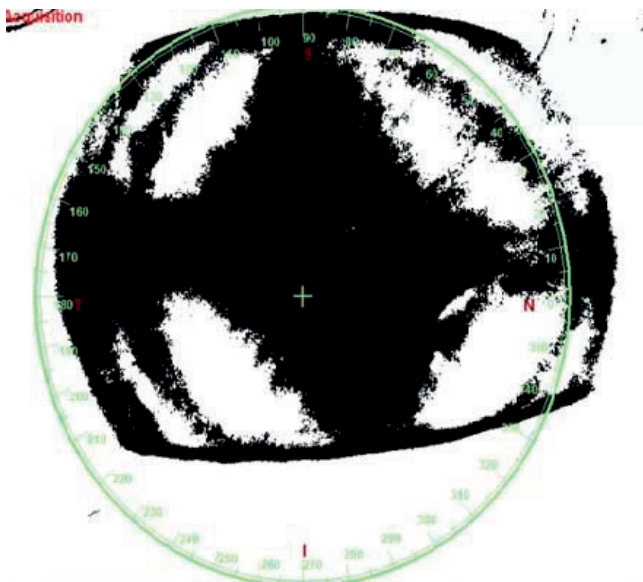


Fig. 1. Lumaxis isogyre acquisition.

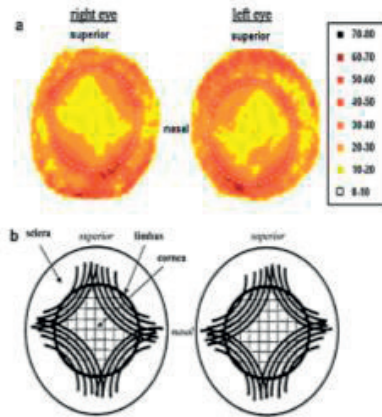


Fig. 2. (a) Contour maps of aligned collagen x-ray scatter from a left/right pair of normal human corneas. Note the high degree of body-line mirror symmetry. (b) Theoretical model of fibrillar arrangement based on (a). Reproduced from Boote *et al.*<sup>30</sup>

## 4. Results

Data obtained by the polarimetric interferometry showed a cross-like pattern (isogyre), shown in Figure 1, which perfectly correlates with the pattern obtained by the x-ray, THG, and SHG analysis (Figs. 2 and 3), confirming that deep stroma lamellae have two preferential alignments along the superior-inferior and nasal-temporal directions. The regular distribution of stromal lamellae allows the stroma to behave as a polarizer, which eliminates the diffractive effect of the light during its journey into the stroma.

## 5. Conclusions

In accordance with the importance of regularity and orientation of stromal lamellae distribution in the corneal refractive process, the importance of polarimetric interferometry as a non-invasive technique to detect such orientation as a consequence of correlation/decorrelation between probe light polarization plane angle and stromal lamellae orientation becomes evident. Information from the cross-like pattern can be used for multiple applications in ophthalmology, such as corneal transplantation and diagnosis of corneal diseases due to stromal pathologies. This new technique represents a unique method to correlate the internal stromal structure and optical properties of the cornea. Recent studies have shown a new, intrinsic axis of the stroma, which can have very important refractive applications in ophthalmology.

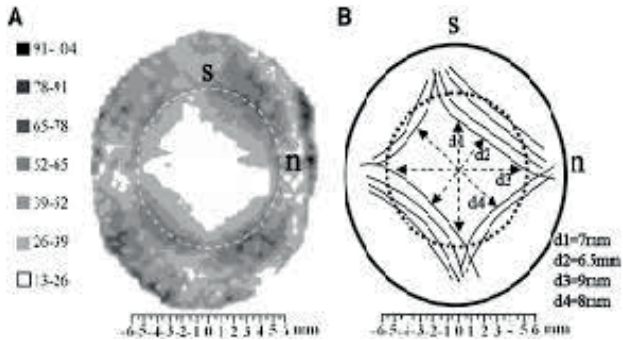


Fig. 3. (a) Contour map of aligned collagen x-ray scatter (a.u.) from a right human cornea. Superior, s, and nasal, n, positions are marked. Broken line denotes the limbus. Note the skewed diamond shape of the scatter contours, which displays mirror symmetry between the left and right eyes. (b) Proposed model of collagen fibril arrangement to explain the shape of the aligned scatter contours. The peripheral, oblique cornea is reinforced by chords of anchoring collagen of scleral origin. Figure modified from Boote *et al.*<sup>31</sup>

## Acknowledgements

Eugenio Lipari is the physicist who has developed this new technique; he is also CEO of Phronema SRL.

## References

1. Pierscionek BK, Popiolek-Masajada A, Kasprzak H. Corneal shape change during accommodation. *Eye*. 2001;15:766-769.
2. Maurice DM. The structure and transparency of the corneal stroma. *Physiol*. 1957; 136:263-286.
3. Hart RW, Farrell RA. Light scattering in the cornea. *J Opt Soc Am*. 1969;59:766-774.
4. Farrell RA, McCally RL, Tatham PER. Wavelength dependencies of light scattering in normal and cold swollen rabbit corneas and their structural implications. *J Physiol*. 1973;233:589-612.
5. Fratzl P, Daxer A. Structural transformation of collagen fibrils in corneal stroma during drying: An x-ray scattering study. *Biophys J*. 1993;64:1210-121.
6. Komai Y, Ushiki T. The three dimensional organization of collagen fibrils in the human cornea and sclera. *Invest Ophthalmol Vis Sci*. 1991;32:2244-2258.
7. Naylor J. Polarized light studies of corneal structure. *Brit. J. Ophthal.*, 1953; 37: 77E.
8. Stanworth A, Naylor EJ. The polarisation optics of the isolated cornea. *Br J Ophthalmol*. 1950;34:201-211.
9. Meek KM, Blamire ST, Elliott GF, Gyi TJ, Nave C. The organization of collagen fibrils in the human corneal stroma: A synchrotron x-ray diffraction study. *Curr Eye Res*. 1987;6:841-846.
10. Benedek GB. Theory of transparency of the eye. *Appl Opt*. 1971;10:459-473.
11. Twersky V. Transparency of pair-related, random distributions of small scatterers, with application to the cornea. *Opt Soc Am*. 1975;65:524-530.

12. Nyquist GW. Rheology of the cornea: Experimental techniques and results. *Exp Eye Res.* 1968;7:183-188.
13. Nash I, Greene P, Foster S. Comparison of mechanical properties of keratoconus and normal cornea. *Exp Eye Res.* 1981;35:413-423.
14. Asejczyk-Widlicka M, Śródka DW, Kasprzak H, Pierscionek BK. Modelling the elastic properties of the anterior eye and their contribution to maintenance of image quality: the role of the limbus. *Eye.* 2006;1-8. Available from: <http://www.nature.com/eye/journal/vaop/ncurrent/pdf/6702464a.pdf>.
15. Asejczyk-Widlicka M, Pierscionek BK. Elasticity and rigidity of the outer coats of the eye. *Br J Ophthalmol.* 2008;92:1415-1418.
16. Wood EA. An Introduction to Optical Crystallography. In: Wood EA. *Crystals and Light.* Princeton, New Jersey: D. Van Nostrand Company, Inc.;1964.
17. Brewster D. On the structure of the crystalline lens in fishes and quadrupeds as ascertained by its action on polarised light. *Phil. Trans R Soc London* 1816;106:311-317.
18. Brewster D. On the development and extinction of regular doubly refracting structures in the crystalline lenses of animals after death. *Phil Trans R Soc London.* 1837;127:253-258.
19. Cope WT, Wolbarsht ML, Yamanashi BS. The corneal polarisation cross. *J Opt Soc Am.* 1978;68:1139-1141.
20. Pierscionek BK, Weale RA. Is there a link between corneal structure and the 'corneal cross'? *Eye.* 1997;11:361-364.
21. Bueno JM, Jaronski JW. Spatially resolved polarization properties for in vitro corneas. *Ophthalmic Physiol Opt.* 2001;21:384-392.
22. Bueno JM, Vargas-Martin F. Measurements of the corneal birefringence with a liquid-crystal imaging polariscope *Appl Opt.* 2002;41:116-124.
23. Jaronski JW, Kasprzak HT. Linear birefringence measurements of the in vitro human. *Ophthalmic Physiol Opt.* 2003;23:361-9.
24. Knighton RW, Huang XR. Linear birefringence of the central human cornea. *Invest Ophthalmol Vis Sci.* 2002;43:82-86.
25. Perutz MF. Polarization dichroism, form birefringence, and molecular orientation in crystalline haemoglobins. *Acta Crystallogr.* 1953;6(11-12):859-864.
26. Bragg WL, Pippard AB. The form birefringence of macromolecules. *Acta Crystallogr.* 1953;6(11-12):865-867.
27. Oldenbourg R, Ruiz T. Birefringence of macromolecules. Wiener's theory revisited, with applications to DNA and tobacco mosaic virus. *Biophys J.* 1989;56(1):195-205.
28. Pierscionek BK, Chan DY. Refractive index gradient of human lenses. *Optom Vis Sci.* 1989 Dec;66(12):822-829.
29. Patent N. WO2014033697; WO2015011692.
30. Meek KM, Boote C. The use of X-ray scattering techniques to quantify the orientation and distribution of collagen in the corneal stroma. *Prog Retin Eye Res.* 2009;28(5):369-392.
31. Boote G, Kamma-Longer CS, Hayes S, et al. Quantification of collagen organization in the peripheral human cornea at micron-scale resolution. *Biophys J.* 2011;101(1):33-42.